



Scaffold-hopping provides both a conceptual and practical route for generating new lead series and chemistries with improved efficacy and pharmacokinetic properties based on known drugs and drug-target interactions.

Classification of scaffold-hopping approaches

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The general goal of drug discovery is to identify novel compounds that are active against a preselected biological target with acceptable pharmacological properties defined by marketed drugs. Scaffold hopping has been widely applied by medicinal chemists to discover equipotent compounds with novel backbones that have improved properties. In this article we classify scaffold hopping into four major categories, namely heterocycle replacements, ring opening or closure, peptidomimetics and topology-based hopping. We review the structural diversity of original and final scaffolds with respect to each category. We discuss the advantages and limitations of small, medium and large-step scaffold hopping. Finally, we summarize software that is frequently used to facilitate different kinds of scaffold-hopping methods.

Introduction

In a modern drug discovery, biologically relevant compounds are usually generated from high-throughput screening (HTS) or virtual screening (VS). For a new target, HTS might be the only way to identify bioactive compounds. However, for targets that are well known, retrieval of active compounds by screening tens of thousands to millions of structurally diverse compounds is neither economical nor efficient. Actually, owing to the limited number of druggable targets [1,2], a large fraction of therapeutically interesting targets are not new and exploration of novel chemistries for these targets could be based on known ligands or ligand–protein complex structures. Historically, many marketed drugs were derived from other known drugs or natural products [3,4]. Thereafter, an important question arises in how to design economically viable drugs based on this knowledge while at the same time maintaining or improving efficacy and pharmacokinetic (PK) profiles of existing therapies by designing novel structural scaffolds (chemotypes).

Scaffold hopping, also known as lead hopping [5,6], is one strategy for discovering structurally novel compounds [7]. Scaffold-hopping methods typically start with known active compounds and end with a novel chemotype by modifying the central core structure of the molecule [3]. Although the concept of scaffold hopping is relatively young [8,9], the strategy has been applied since the beginning of drug discovery. It has not only applied to jumpstart a project with known

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ligands, scaffold hopping is also widely used in lead optimization approaches [10,11]. Because many compounds in corporate libraries are failed compounds with poor physicochemical and PK properties, the hit molecules from HTS can inherit these unfavorable properties. These properties depend on how mature the compounds are in the drug discovery pipeline. Sometimes, modification of side chains is sufficient to overcome the undesirable properties associated with the parent molecule, whereas at other times, the core structure or the scaffold of the parent molecule has to be modified.

In the past decade, published research works on scaffold hopping have increased exponentially [10,11], yet limited effort has been made to characterize the frequently used scaffold-hopping methods. In this review, scaffold hopping is classified into four major categories – heterocycle replacements, ring opening or closure, peptidomimetics and topology-based hopping. The structural diversity of original and final scaffolds with respect to each category will be reviewed. The advantages and limitations of small, medium and large-step scaffold hopping will also be discussed. The small step hops, represented by swapping carbon and nitrogen atoms in an aromatic ring or replacing carbon with other heteroatoms in a ring, results in a low degree of structural novelty. By contrast topology-based hops always lead to a high degree of novelty. Significantly more examples of small to medium step scaffold hopping are found in publications. This illustrates the tradeoff between degree of structural novelty and the success rate of achieving comparable biological activities [3]. There are many methodologies that enable the generation of scaffold hops. These include use of 2D fingerprints and 3D pharmacophores in the context of VS. These ‘enabling’ technologies are out of the scope of this article. The interested reader can find illuminating discussions on these topics in the following references [5,12–18].

History

The concept of scaffold hopping was introduced in 1999 by Schneider *et al.*, as a technique to identify isofunctional molecular structures with significant different molecular backbones [9]. The simple definition emphasized two key components of scaffold hopping: different core structures and similar biological activities of the new compounds relative to the parent compounds. The two requirements seem to conflict with the similarity property principle, which states that compounds with similar chemical structures usually possess similar physicochemical properties and biological activities [19]. The principle does not exclude the possibility of structurally diverse compounds from binding to a same target. The similarity property principle is generally held because ligands that can fit in the same pocket should share certain structural similarity (i.e. similar shape and similar electropotential surface) although the ligands might belong to different chemotypes. Owing to the fact that the repulsive penalty in the Lennard-Jones potential is extremely sensitive to the interatomic distances, the landscape of activity is far from linear; the addition or removal of a small methyl group might result in large changes in biological activity [20], whereas the similarity metrics are insufficient in reflecting this nonlinearity. In addition, the flexibility of both proteins and small molecules further complicates the relationship between similarity and activity. Nevertheless, the similarity property principle is still the pillar of modern drug discovery, such

as structure–activity relationships (SAR), and scaffold hopping is not an exception of this principle.

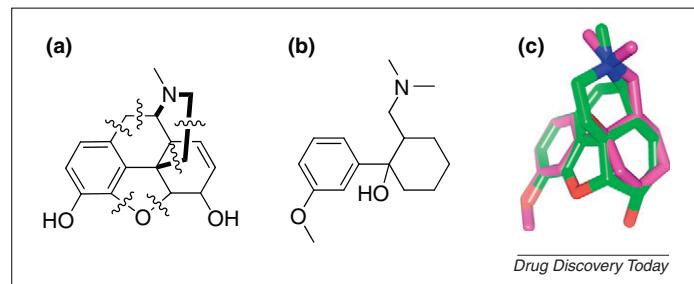
Historically, a large fraction of marketed drugs are derived from natural products, natural hormones and other drugs through scaffold modification [4]. Revisiting these successful examples and other newly published examples supplies useful guidance for medicinal chemists to create novel chemical entities based on known bioactive molecules.

Morphine and tramadol

Opium, one of the earliest known drugs, has been used for over a thousand years for relief of pain. Morphine, the major component of opium, is a potent analgesic, but its medical use is limited by its addictive potential. Morphine acts on the μ -opioid receptor to increase tolerance to painful stimuli. Besides its addictive liability, morphine has other adverse side effects, including nausea, vomiting and respiratory depression. Morphine is a rigid ‘T’ shaped molecule (Fig. 1a). By breaking six ring bonds and opening up three fused rings, the new drug tramadol is more flexible, resulting in reduced potency and reduced side effects (Fig. 1b). The 2D structures of morphine and tramadol are very different, but 3D superposition of both molecules, as calculated by using the Flexible Alignment program in Molecular Operating Environment (MOE) [21], demonstrates that the key pharmacophore features are conserved. Fig. 1c shows the shared spatial position of the positively charged tertiary amine, the aromatic ring and the hydroxyl group attached to phenyl ring (the methoxyl group in tramadol is demethylized by CYP2D6). Although tramadol is only one-tenth of the potency of morphine, it is almost completely absorbed after oral administration, and lasts for up to six hours. The transformation from morphine to tramadol by ring opening is one of the earliest examples of scaffold hopping [4].

Antihistamines

Histamine is an organic nitrogen compound derived from decarboxylation of the amino acid histidine [22]. It has multiple physiological functions, including triggering the inflammatory response and regulating immune response, sleep and allergies [22]. The classical antihistamines possess two aromatic rings joined to one carbon or nitrogen atom and together with one positive charge center. This is represented by the structure of pheniramine (Fig. 2a). Pheniramine, also known as Avil, is an antihistamine used to treat allergic conditions, such as hay fever or urticaria [23]. It competes with histamine for histamine H1-receptor sites, to reduce the intensity of allergic reactions and tissue injury response involving histamine release. An analog of pheniramine called cyproheptadine, has significantly improved binding affinity against the H1-receptor. This was achieved by locking both aromatic rings of pheniramine to the active conformation through ring closure, and by introducing the piperidine ring to further reduce the flexibility of the molecule (Fig. 2b). This rigidified molecule is also better absorbed. In addition, the structural changes achieve other medical benefits in the prophylaxis of migraine, because cyproheptadine can antagonize the serotonin or 5-hydroxytryptamine (5-HT) type 2 receptor. Isosteric replacement of one phenyl ring in cyproheptadine with thiophene produces pizotifen (Fig. 2c), which proves to be a better medicine for the treatment of migraine [24]. Azatadine (Fig. 2d) was developed

**FIGURE 1**

Structures of pain-killing drugs: (a) morphine, (b) tramadol and (c) 3D superposition of (a) in green and (b) in magenta.

by the Schering-Plough Corporation (<http://www.merck.com/index.html>) as a typical potent sedating antihistamine [25]. It is formed from cyproheptadine by replacing one phenyl ring with pyrimidine that improves the solubility of the molecule. The 2D structures are different, but 3D superposition shows that the pharmacophore orientation is similar [e.g. the spatial position of the basic nitrogen and the two aromatic rings overlap (Fig. 2e,f)].

These antihistamine examples show that small changes in molecular structures can result in different activity profiles, thus different medical uses. It also clearly demonstrates that reduction of molecular flexibility can increase the potency of molecules, presumably by reducing entropy loss upon binding to the targets.

Classification of scaffold-hopping approaches

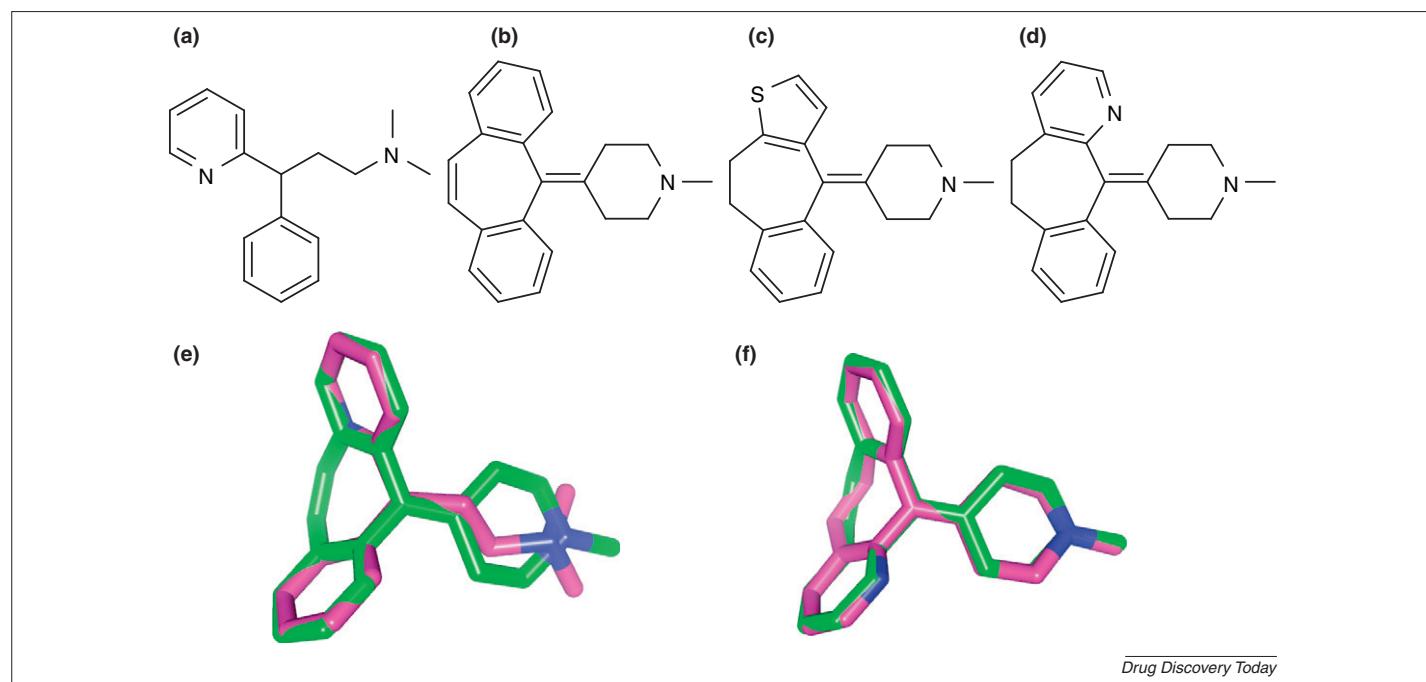
According to the definition of scaffold hopping, derivatives obtained from the parent compounds have novel core structures. The question is how different the derivative molecules must be

from their parents in order for the evolution to be classified as scaffold hopping. In other words, how novel is novel? Boehm *et al.* classified two scaffolds as different if they were synthesized using different synthetic routines, no matter how small the change might be [3]. This statement has been proven true in many cases where the chemical structures are closely related but different patents can be claimed, or different new drug applications can be approved by the U.S. Food and Drug Administration (U.S. FDA). For example, the major structural variation between the two phosphodiesterase enzyme type 5 (PDE5) inhibitors sildenafil and vardenafil is the swap of a carbon atom and a nitrogen atom in the 5–6 fused ring (Fig. 3a,b) but the difference is enough for the two molecules to be covered by different patents [26]. The two cyclooxygenase 2 (COX-2) inhibitors rofecoxib (VioxxTM) and valdecoxib (BextraTM) differ by only the five-member hetero rings connecting the two phenyl rings (Fig. 3c,d), yet they were sold by Merck and Pharmacia/Pfizer (<http://www.pfizer.com/home/>) separately [27].

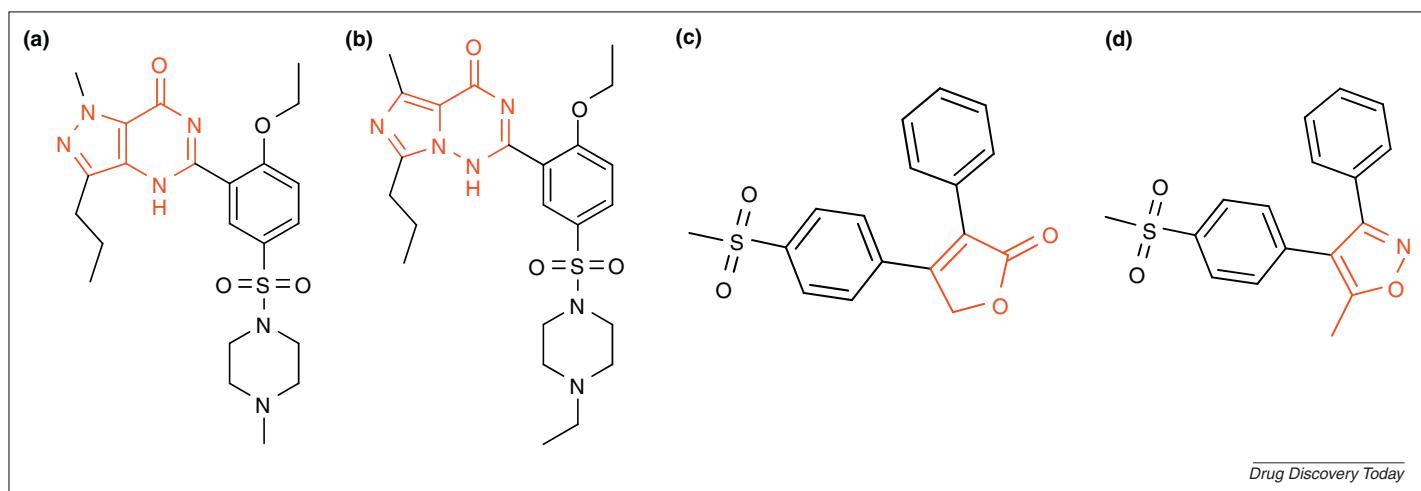
We rationalize the concept of scaffold hopping by focusing on the degree of change associated with the original parent molecule. Minor modifications like replacing or swapping carbon and heteroatoms in a backbone ring, are classified as a 1° hop. More extensive ring opening and closures is classified as a 2° hop. Replacement of peptide backbones with nonpeptic moieties falls into the category of a 3° hop. Finally a complete new chemical backbone that only retains interactions is characterized as a 4° hop.

1° hop: heterocycle replacement

The heterocycles functioning as cores of drug molecules usually provide multiple vectors projecting to different directions. Replacing the C = carbon, N = nitrogen, O = oxygen and S = sulfur

**FIGURE 2**

Structures of antihistamine drugs (a) pheniramine, (b) cyproheptadine, (c) pizotifen, (d) azatadine, (e) superposition of drugs (a) in magenta and (b) in green and (f) superposition of drug (b) in green and (d) in magenta.



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FIGURE 3

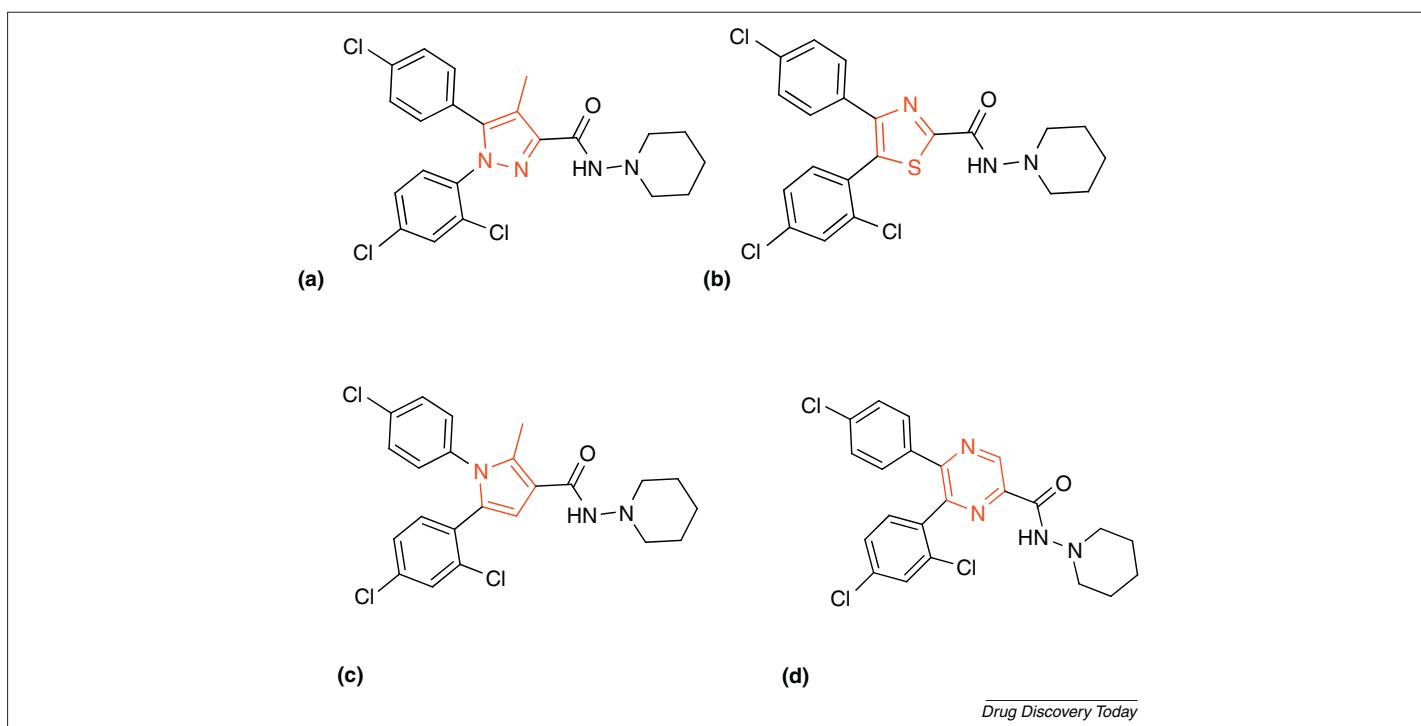
Structures of PDE5 inhibitors (a) sildenafil, (b) vardenafil and cyclooxygenase (COX-2) inhibitors, (c) rofecoxib and (d) valdecoxib. Abbreviation: PDE5: phosphodiester enzyme type 5.

atoms in a heterocycle, while maintaining the outreaching vectors can result in novel scaffolds. Improved binding affinity is likely to be achieved if the heterocycle is directly involved in interactions with the target protein.

Cannabinoid 1 receptor inhibitors

Rimonabant (AcompliaTM) is an anorectic antiobesity drug produced and marketed in Europe by Sanofi-Aventis (<http://en.sanofi.com/>). Its inverse agonist effect on the cannabinoid 1 (CB1) receptor causes a reduction in appetite. Rimonabant was the first selective CB1 receptor antagonist to be approved for use in

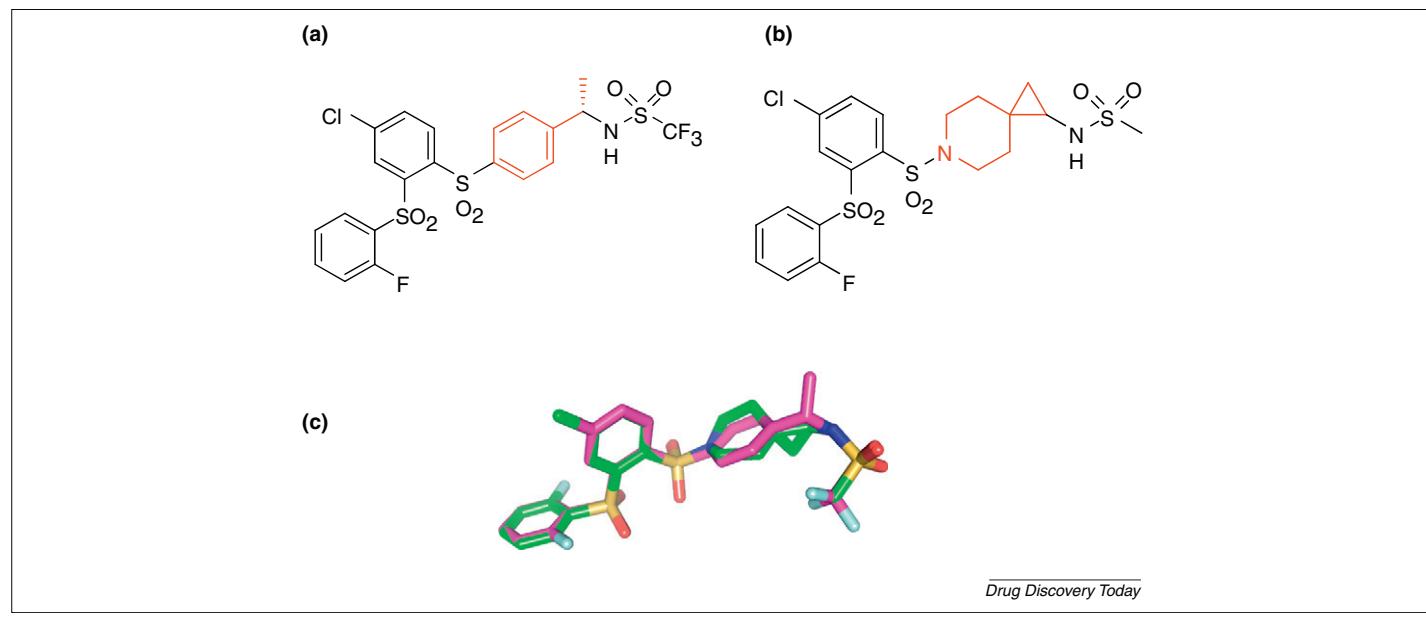
humans. However, the antiobesity drug failed to win approval from the U.S. FDA to enter the US market, due to safety concerns. Bostroem's group at AstraZeneca (<http://www.astazeneca.com/Home>) initiated a scaffold-hopping approach, attempting to discover novel CB1 antagonists with improved physicochemical and drug metabolism and pharmacokinetics (DMPK) properties [28]. They tried to replace the methylpyrazole core in rimonabant with a range of five- and six-member rings, including thiazoles, pyrroles and pyrazines (Fig. 4) [29,30]. The newly designed compounds were ranked by ease of synthesis and shape similarity against rimonabant, as computed with Rapid Overlay of Chemical



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FIGURE 4

Structures of the cannabinoid 1 (CB1) antagonists (a) rimonabant, (b) thiazole derivative, (c) pyrrole derivative, and (d) pyrazine derivative.

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Structures of a triaryl bis-sulfone CB2 receptor inhibitor (a) and its biaryl analog (b). The superposition of both structures (c) (molecule (a) in magenta and (b) in green) was calculated by using the Flexible Alignment program in MOE [21]. Abbreviations: CB2: cannabinoid 2; MOE: Molecular Operating Environment.

Structures (ROCS) [31]. All three new scaffolds resulted in novel classes of CB1 receptor antagonists, but their safety profiles have not been fully examined [28].

CB2 inhibitors

Sharing 44% sequence similarity with the CB1 receptor, the CB2 receptor is expressed primarily in cells of the immune system [32]. Modulation of the immune system might be realized through antagonists and inverse agonists of the CB2 receptor. Merck scientists [33] discovered a potent and selective triaryl bis-sulfone CB2 inhibitor a few years ago (Fig. 5a). In a recent backup program, they attempted to remove some unfavorable activities, such as calcium channel blockage and cytochrome P450 2C9 inhibition, associated with the triaryl compound. To achieve this goal, they searched for less hydrophobic analogs. By replacing the central phenyl ring in the triaryl compound (Fig. 5a) with spiracyclopropyl piperidine (Fig. 5b), the biaryl derivative demonstrated the same potency against CB2 and selectivity against CB1. This is indicated by their superimposed structures as shown in Fig. 5c. Furthermore, the rat calcium channel affinity was reduced from 0.5 μ M to 8 μ M, and the 2C9 activity reduced from 3.5 μ M to 30 μ M. The replacement of the third aromatic ring with the saturated ring system also makes the molecule more drug-like [34,35], with the calculated log *P* value dropping from 2.81 to 1.48 [36].

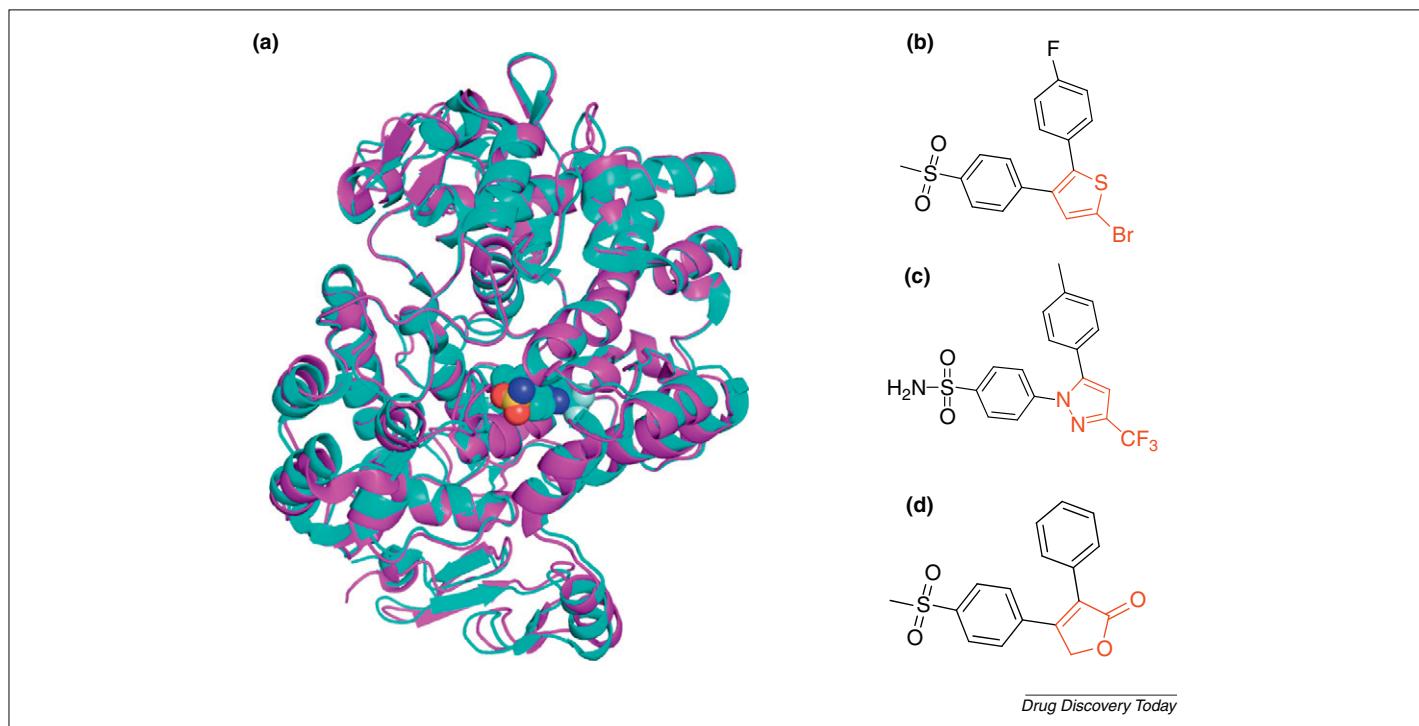
Cyclooxygenase-1 and cyclooxygenase-2 inhibitors

Nonsteroidal anti-inflammatory drugs (NSAIDs) function by inhibiting the enzyme cyclooxygenase (COX), which catalyses the biosynthesis of prostaglandins (PGs) from arachidonic acid (AA). There are two enzymes in humans that catalyze the first

step in the biosynthesis of PGs, namely COX-1 and COX-2. Although catalyzing the same reaction, COX-1 and COX-2 are different in sequence (~60% identity), tissue distribution and physiological function. The COX-1 isozyme has a role in gastroprotection and vascular homeostasis, whereas the COX-2 isozyme is mainly involved in inflammatory processes [37,38]. Selective inhibition of the COX-2 isozyme could circumvent the adverse ulcerogenic effects associated with classical NSAIDs, such as aspirin and ibuprofen [39]. Although COX-1 and COX-2 only share 60% sequence homology, the protein backbones, especially the ligand-binding sites, are very similar to each other (Fig. 6a) [40,41]. By contrast, the subtle structural differences at the ligand-binding sites are sufficient to generate COX-2 selective inhibitors [42].

The first COX-2 selective inhibitor, DuP 697 (Fig. 6b), was discovered in 1990 [43]. The structure of DuP 697 was the template for the design of the diarylheterocyclic family of selective COX-2 inhibitors. These include later marketed drugs celecoxib and coxib (Fig. 6c,d). However, DuP 697 failed to reach the market due to safety issues [39]. Rofecoxib, also known as Vioxx, was withdrawn from the market owing to concerns about increased risk of heart attack and stroke associated with long-term and high-dosage use, whereas its close analog, celecoxib, is still in use for the treatment of osteoarthritis, rheumatoid arthritis, acute pain, painful menstruation and menstrual symptoms.

The three diarylheterocyclic COX-2 selective inhibitors differ from each other mainly in the backbone heterocyclic rings (Fig. 6b–d). Their activity levels against COX-2 are comparable [39] but their pharmacology is totally different. It is not yet fully understood whether the relative selectivity levels or the kinetic behavior of inhibition causes the differences [40,44], but the

**FIGURE 6**

(a) Overlay of X-ray crystal structures of COX-1 in magenta (PDB id: 3KK6) and COX-2 in cyan (PDB id: 3LN1) in complex with celecoxib, and structures of diarylheterocyclic COX-2 selective inhibitors (b) DuP 697, (c) celecoxib and (d) rofecoxib. Abbreviation: COX-1: cyclooxygenase 1.

effects of heterocyclic replacement on the PK and pharmacodynamic behavior of a compound are clearly demonstrated in this example. This example shows that the impact of 1° scaffold hopping on the mechanism of action (MOA) of a drug molecule is not always predictable. However, 1° scaffold hopping is a good strategy to improve the success rate in drug discovery.

Software

For 1° hopping, the software MORPH is a useful tool for generating novel aromatic rings systematically without changing the coordinates of the ring atoms [45]. MORPH can generate new molecules by altering individual rings and fused ring systems, while at the same time maintaining user-defined constraints. To overcome the potential issues regarding synthetic feasibility and chemical stability associated with creating novel aromatic systems systematically [46], natural products and marketed drugs can serve as templates for generating a pool of drug-like scaffolds [47]. Another molecular design software package, Recore [48], is also based on this kind of scaffold library generation. A comprehensive list of software used in scaffold hopping is presented by Brown *et al.* [10,16].

2° hop: ring opening and closure: pseudo ring structures

Most drug-like molecules contain at least one ring system, so ring opening and ring closure are two immediate strategies to create novel scaffolds. Because molecular flexibility contributes greatly not only to the entropic component of the binding free energy but also to membrane penetration and absorption [49], ring opening and closure are useful strategies for improving the drug-like

properties of molecules. Ring opening and closure manipulate the flexibility of a molecule by controlling the total number of free rotatable bonds. This can be accomplished in a variety of ways.

Ring closure

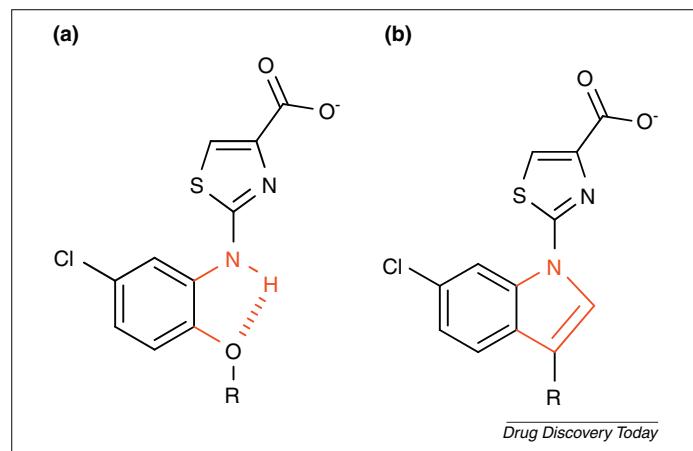
Intramolecular hydrogen bonds (HBs) usually offer direct hints on where to close a ring, as shown in following examples. Intrigued by the potential intramolecular HB between the *o*-alkoxy group and biaryl NH (Fig. 7a), a GlaxoSmithKline (<http://www.gsk.com/>) group synthesized a series of indole compounds (Fig. 7b) as PG EP1 receptor antagonists [50]. The ring closure design successfully locked the molecule into a bioactive conformation. One of the resulting indole compounds (Fig. 7b), where R was *iso*-butyl, showed low nM and sub-nM activities in binding and functional EP1 antagonist assays [50].

Other frequently used ring closure ideas include converting an alkyl chain to cyclohexane, piperazine or piperidine [51], converting an *o*-hydroxylbenzoyl group to quinazoline [52], and converting an arylamine or arylamide to a fused ring system [53].

It is worth noting that reduction in entropy loss due to binding might be limited, because the intramolecular HBs of the parent compounds have already reduced molecular flexibility.

Ring opening

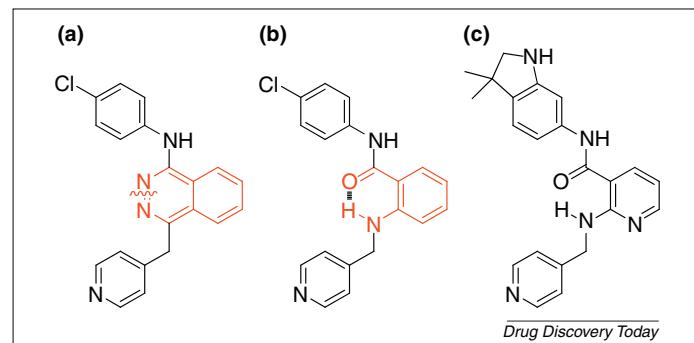
Although ring closure has a positive impact on the binding free energy, it does so by producing potentially negative impacts on solubility and other ADME properties [34]. To overcome the adverse effects of too many rings in a molecule, medicinal chemists might practice ring opening, to enhance the drug-likeness of molecules.

**FIGURE 7**

Prostaglandin EP1 receptor antagonists: (a) biaryl amine series and (b) indole series.

Pyridopyrimidinone is a typical moiety of protein kinase inhibitors. PD166285 (Fig. 8a), a 6-aryl substituted pyridopyrimidinone, is a broad-spectrum tyrosine kinase inhibitor [54]. In an attempt to design novel tyrosine kinase inhibitors using PD166285 as a template, Furet *et al.* opened up the pyrimidine ring and moved the nitrogen atom in position 1 of the pyrimidine ring to position 5 to form a pseudo six-member ring with the adjacent urea through intramolecular hydrogen bonding (Fig. 8b) [55]. Because the urea in pyrimidinyl urea did not adopt the low energy extended conformation, *ab initio* calculations and data mining were carried out to confirm that the pseudo cyclic conformation was favorable. The pseudo ring design concept was further supported by the assay results; the pyrimidinyl urea compound (Fig. 8b) exhibited submicromolar inhibition against several tyrosine kinases, such as c-Src, EGFR and c-Abl [55]. Although the physicochemical properties of the pseudo ring compound and its parent bicyclic compound were not mentioned in the paper, the calculated log *P* value of the urea derivative is 1 log unit lower than its parent compound [36].

In another approach involving novel antiangiogenic agents, a Novartis (<http://www.novartis.com/>) team opened the phthalazine ring in a known dual KDR and Flt-1 inhibitor PTK787/ZK222584 (Fig. 9a) [56]. The anthranilic amide moiety is ready to form a six-member pseudo ring through intramolecular hydrogen bonding (Fig. 9b). Ring opening resulted in minimal changes on activity and selectivity [56]. A close analog of this compound, Motesanib/AMG 706 (Fig. 9c) was successfully transitioned to clinical trials [57]. This example demonstrates that ring opening

**FIGURE 9**

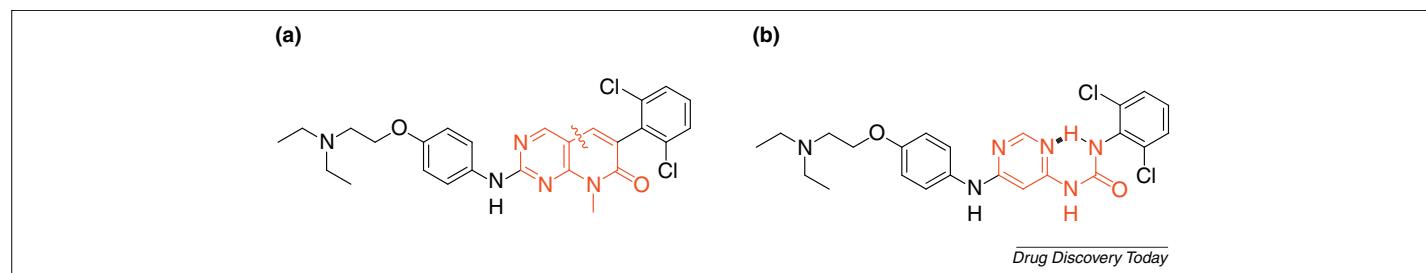
Structures of antiangiogenic agents: (a) phthalazine PTK787/ZK222584, (b) its anthranilic amide analog and (c) Motesanib/AMG 706.

was not only a useful strategy to generate new chemical classes from templates but also a promising method to improve drug-like properties.

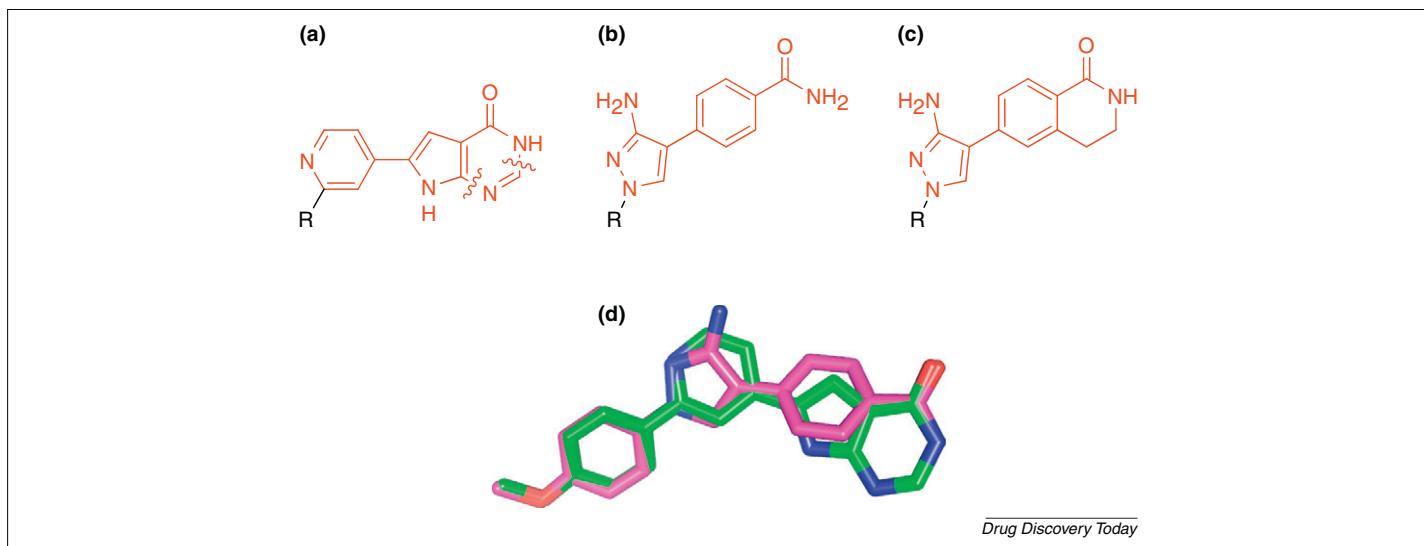
Ring opening and closure

Ring opening and ring closure can be applied simultaneously on the same molecule, which might cause ring shift or ring migration [58]. Mitogen-activated protein (MAP) kinase-activated protein kinase 2 (MK2) has a crucial role in signaling and synthesis of tumor necrosis factor α (TNF- α). Potent and selective MK2 inhibitors can potentially be developed into anticancer drugs. To discover novel MK2 inhibitors, Velicky's group opened up the pyrimidinone ring of the parent pyrrolo-pyrimidone structure (Fig. 10a) while maintaining the attached amide group, and then swapped the five- and six-member rings [59]. The resulting compound (Fig. 10b) was fourfold less potent than its parent compound. Reconnecting the amide group to the phenyl ring to reduce the flexibility of the amide boosted the affinity a factor of 25. The final dihydroisoquinolinone compound (Fig. 10c) showed 84 nM potency against MK2 [59]. Although there is significant structural variation between the original and final compounds, both scaffolds overlaid well (Fig. 10d). Introduction of the two sp^3 carbon atoms in ring closure increased the saturation level of the molecule, thus enhancing the possibility of the compound becoming a drug [60].

Another special ring closure yields macrocyclic molecules. Macrocycles here refer to those molecules with rings containing nine or more atoms. This kind of ring closure is used to constrain the conformation [61–64]. Because the whole molecule is changed from a linear structure to a cyclic structure, macrocyclic ring closure is a special kind of scaffold hopping.

**FIGURE 8**

Structures of tyrosine kinase inhibitors: (a) pyridopyrimidinone PD 166285 and (b) its urea derivative.



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FIGURE 10

Structures of MK2 inhibitors: (a) pyrrolo-pyrimidone template, (b) amide analog, (c) dihydroisoquinolinone derivative and (d) the overlay of (a) in green and (b) in magenta. Abbreviation: MK2: mitogen-activated protein (MAP) kinase-activated protein kinase 2.

In summary, ring closure can potentially lock a flexible molecule into its bioactive conformation, thereby reducing the entropy penalty upon interacting with the target protein. By contrast, too many rings, especially aromatic rings, in one molecule tend to reduce the drug-likeness of the molecule [60]. Introduction of saturated ring systems can both suppress the molecular flexibility and maintain drug-likeness, but the synthetic feasibility will inevitably suffer from newly introduced chiral centers or spiro-like structures [65].

Software

There is no particular software available to generate ring opening or closure design ideas. Because variation of the ring system in a molecule is usually associated with conformational changes, the Cambridge Structural Database (CSD) [66,67] is a valuable source for low-energy molecular conformations that can be used to validate design concepts [68].

3° hop: pseudopeptides and peptidomimetics

Biologically active endogenous peptides, such as peptide hormones, growth factors and neuropeptides have a vital biological function in our bodies. Imbalance of these peptides can cause different human diseases, including diabetes, cancer, osteoporosis and endometriosis [69]. The development of peptides into clinically useful drugs is largely hampered by their poor metabolic stability and low bioavailability [69]. Design of small molecules to mimic the structural features of peptides using active peptide conformations as templates has shown promising results for some challenging targets [70,71]. The application has been extended to targets involved in protein–protein interactions, where small molecules are designed to mimic the interacting moieties of proteins. The major goal of peptide-based drug discovery is to reduce the peptide character to enhance the resistance to proteolysis, while maintaining the key chemical features for molecular recognition. Scaffold hopping is a typical method used to carry out the peptide to small molecule transition.

Secondary structures such as α -helices [72–74], β -sheets [75] and β/γ -turns [76] are frequently observed at the interfaces of peptide–protein and protein–protein interacting partners. Synthetic structures have been designed to mimic these secondary structures [77–80]. In these designs it is important that the synthetic scaffolds position the side chains consistently with the helical and turn structures. Maintaining the backbone hydrogen-bonding interactions is the major task for β -sheet mimetics. These strategies have been reviewed elsewhere [71], so the focus of this article will be placed on the scaffold-hopping designs where derivative molecules are structurally similar to their parents.

Triggering apoptosis

Apoptosis, or programmed cell death, has a major role in maintaining homeostasis and removal of damaged or malignant cells [81]. Imbalances in apoptosis pathways are linked to several therapeutically important disease areas, including oncology, cardiovascular diseases and neurodegenerative diseases [82–85]. The second mitochondria-derived activator of caspases (Smac) interacts with XIAP X-linked inhibitor of apoptosis (XIAP) by inserting its N-terminal sequence, AVPI (ALA-VAL-PRO-ILE) (Fig. 11a) into the XIAP–caspase-9 interaction pocket, thus releasing caspase-9 and causing cell death. Modified peptides and peptide mimetics have been designed to compete with AVPI/Smac to cause apoptosis. Wist *et al.* replaced one peptide bond with an oxazole ring (Fig. 11b), aiming at enhancing drug-likeness by reducing the peptide character [78]. The resulting Smac mimics, AoxSPF, AoxSPW, AoxSPY and AoxSPI, could bind to the Baculovirus Inhibitor of apoptosis protein Repeat 3 (BIR3) domain of XIAP with much lower binding affinity. AVPI interacts with BIR3-XIAP mainly through backbone hydrogen bonding, forming an antiparallel β -sheet structure. The oxazole replacement changed the hydrogen bonding features of both carbonyl and amine in the peptide bond (Fig. 13), resulting in the loss of key backbone interactions. Indeed, the crystal structure of AoxSPW bound to BIR3-XIAP indicated that the compound formed two fewer HBs with the protein [78].

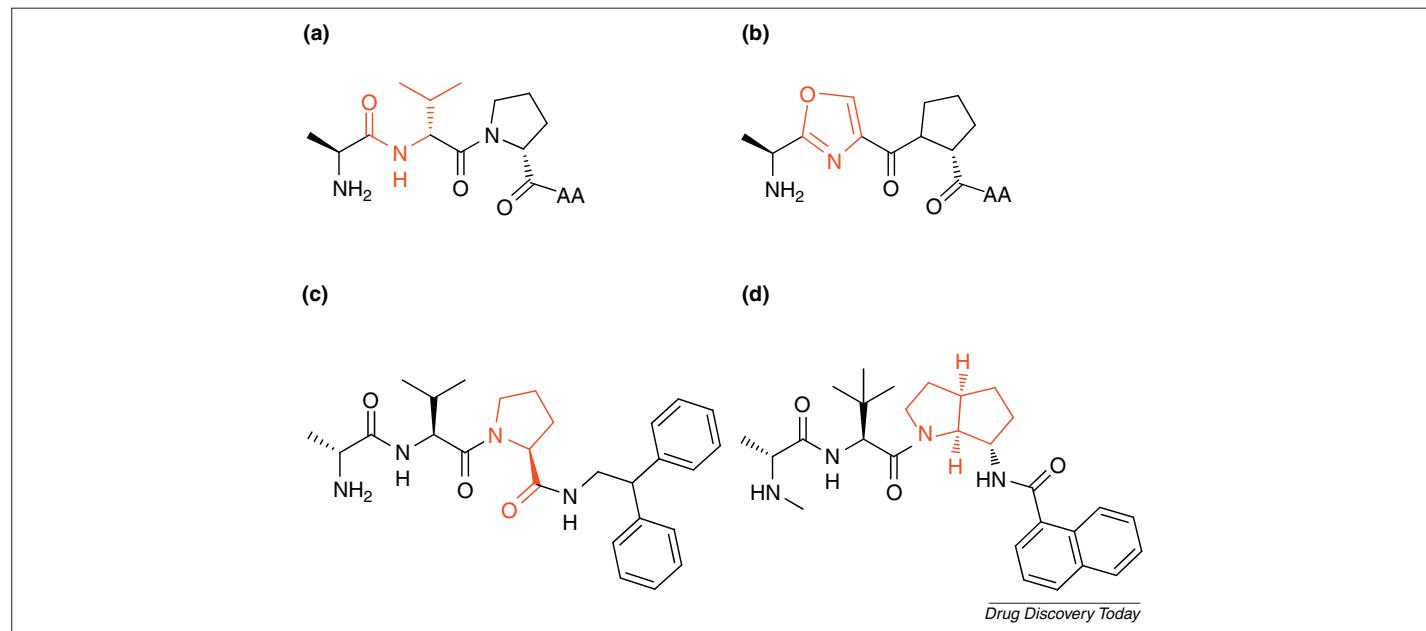


FIGURE 11

Structures of (a) Smac N-terminal tetrapeptide AVPI, (b) an oxazole derivative, (c) modified Smac tetrapeptide and (d) an azabicyclooctane analog. Smac: second mitochondria-derived activator of caspases.

Cohen's group [86] used the software CAVEAT [87] to generate design ideas for small molecules using the crystal structure of a modified tetrapeptide, AVP-2,2-diphenylamine (Fig. 11c), as a template [88]. The original CAVEAT hits were identified as either synthetically infeasible or chemically unstable, but the bicyclic motif inspired the authors to manually search the literature for similar scaffolds. The resulting azabicycloocatane compound (Fig. 11d) demonstrated a better docking score than the tetrapeptide (Fig. 11a), and high binding affinities against XIAP, ML-IAP and c-IAP with K_i values of 140, 38 and 33 nm, respectively [86]. Replacing the side chain of Val with *t*-butyl, and PRO with azabicycloocatane, improved the PK properties and bioavailability of the compound [86]. Similar bicyclic scaffolds were observed in design of the inhibitors of prolyl oligopeptidase (POP), a target for the treatment of neurodegenerative and psychiatric diseases [89].

Angiotensin II

The octapeptide hormone angiotensin II (Ang II) is involved in a range of physiological activities, such as vasoconstriction, aldosterone release, cell differentiation and tissue repair, through interaction with angiotensin 1 and 2 (AT1 and AT2) receptors [90,91]. Ang II, given by the sequence DRVYIHPF (Fig. 12a), is believed to adopt a turn structure centered at Tyr at position 4 while activating the AT1 receptor [92–94]. Several β -turn and γ -turn mimetics have been designed as Ang II receptor ligands [95–97]. By replacing the central residues Tyr at position 4 and Ile at position 5 with a benzodiazepine (Fig. 12b), a well-known β -turn mimetic scaffold [98], the new pseudopeptide exhibited high binding affinities against both AT1 and AT2 receptors with K_i values of 14.9 nm and 1.8 nm, respectively [97]. The benzodiazepine-based β -turn design can position key residues of Ang II into locations similar to that of γ -turn mimetics [97,99]. The metabolic stability of the

pseudopeptides is always a concern owing to the remaining peptide bonds in the molecules, but it was not discussed in the paper.

Software

Recore [48] and CAVEAT [87] are useful for designing suitable scaffolds to replace parts of peptides. Pharmacophore modeling packages by Chemical Computing Group [100], Accelrys [101] and Schrodinger [102] are also widely applied in peptidomimetic design [70,103,104].

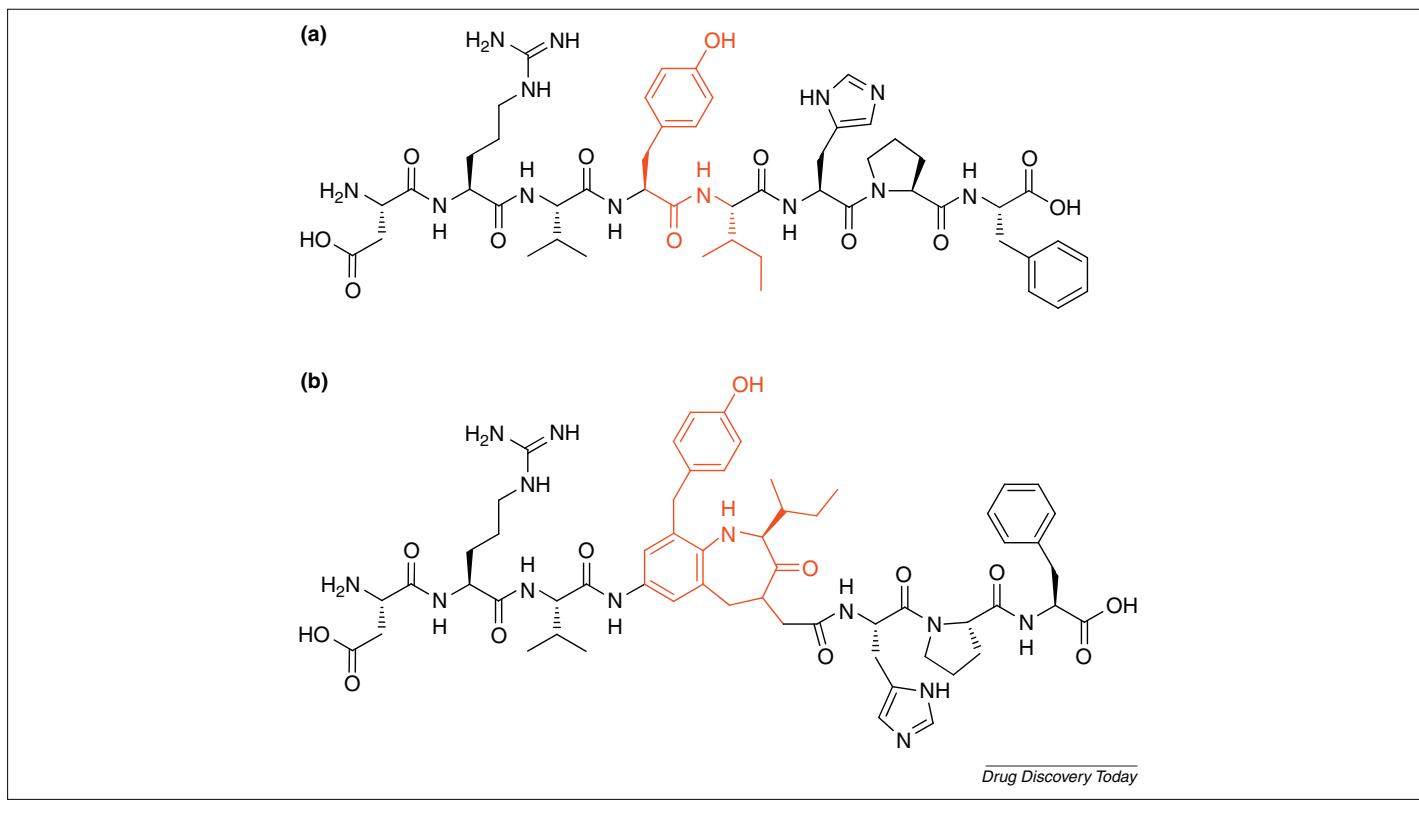
4° hop: topology/shape-based scaffold hopping

Top. topology/shape-based scaffold hopping

Successful stories of topology/shape-based scaffold hopping are rare in the literature. One possible reason is that many attempts have been made, but most failed and thus not published. Another possibility is when the new chemotype is significantly different from its template, scientists might consider the process as VS, rather than scaffold hopping. This type of scaffold hopping can be generated using VS, as demonstrated in the following examples, but we wish to retain the distinction in that VS is a technology that enables scaffold hopping. Ultimately, scaffold hopping focuses on discovering novel core structures, usually ignoring potential conflicts between side chains and targets, whereas VS aims at whole molecules as hits.

Lipoxygenase inhibitors

Schneider and coworkers identified a novel 5-lipoxygenase (5-LO) by similarity searching of a natural product collection and natural product-derived combinatorial libraries. The scaffold-hopping approach, enabled by similarity search, was performed with topological pharmacophore models derived from 43 known 5-LO inhibitors [105]. The scaffold of the best hit was not represented in any of the known inhibitors, and the overall structure was significantly different from the query molecules.

**FIGURE 12**

The structures of (a) Ang II (DRVYIHPF) and (b) benzodiazepine-based β -turn mimetic.

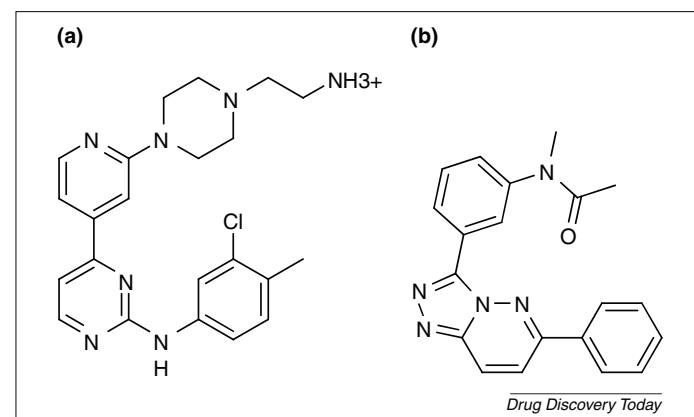
Zipa-FtsZ inhibitors

To pursue novel antibiotics effective against resistant mutants, a Wyeth (<http://www.pfizer.com/welcome/>) team chose to interrupt cell wall biosynthesis by targeting bacterial ZipA-FtsZ protein–protein interactions [106–108]. The template they chose was a weak HTS hit (Fig. 13a), possessing potential toxicity and intellectual property (IP) issues. The use of a shape-based ROCS search with the compound in Fig. 13a as template led to the discovery of novel scaffolds with low 2D similarity against their template. This implies that these ROCS-identified scaffolds could not be found by 2D search methods. The ROCS hits were less potent, but they did not show toxicity and IP issues associated with pyridylpyrimidine core of molecule shown in Fig. 13a [108]. One of the ROCS hits (Fig. 13b) was cocrystallized with ZipA. The binding geometry of the compound shown in Fig. 13b showed similarities to the geometry of template in Fig. 13a used in the ROCS search.

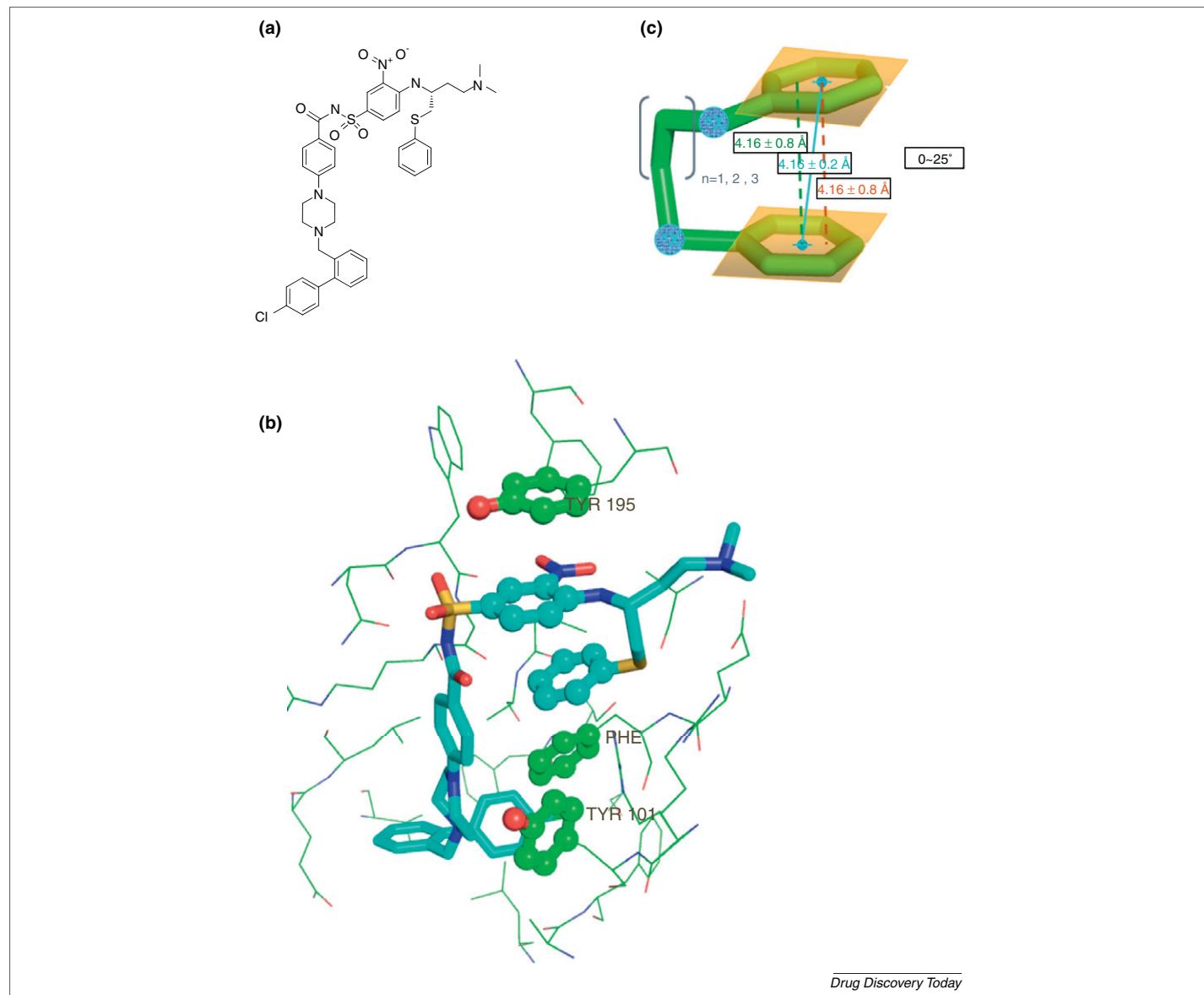
B-cell lymphoma-extra large inhibitors

Sun [109] at Roche (<http://www.roche.com/index.htm>) discovered a new series of B-cell lymphoma-extra large (BCL-xL) inhibitors by using the published structure of ABT-737 (Fig. 14a) as a template [110]. They attributed the tight binding between ABT-737 and BCL-xL to the π – π stacking network formed between the ligand and the protein (Fig. 14b), and attempted to identify a molecular scaffold to mimic the π – π stacking topology. A CSD [67] query was created to capture the key topological requirements, including the angle between the two aromatic planes and the distance between the centroids of both aromatic planes

(Fig. 14c). Searching nearly a half-million crystal structures in CSD led to a novel scaffold mimicking the π – π stacking feature in ABT-737 (Fig. 15a). Taking advantage of transferable SAR, ABT-737 side chains were added to the novel scaffold to achieve comparable binding benefits as its parent template. Finally, with less than a few dozen compounds synthesized, the Roche scientists discovered a series of BCL-xL inhibitors, among which the most active inhibitor (Fig. 15b) was reported equipotent to ABT-737, but more than 200-daltons smaller and much less hydrophobic.

**FIGURE 13**

Structures of ZipA-FtsZ inhibitors: (a) pyridylpyrimidine template from HTS and (b) ROCS-identified hit. Abbreviations: HTS: high-throughput screening; ROCS: Rapid Overlay of Chemical Structures.



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FIGURE 14

(a) Structure of BCL-xL inhibitor ABT-737, (b) the ligand-binding site of BCL-xL illustrating the $\pi-\pi$ stacking network formed between the ligand ABT-737 (cyan) and the protein BCL-xL (green) and (c) the CSD query for reproduction of the $\pi-\pi$ stacking. Abbreviations: BCL-xL: B-cell lymphoma-extra large; CSD: Cambridge Structural Database.

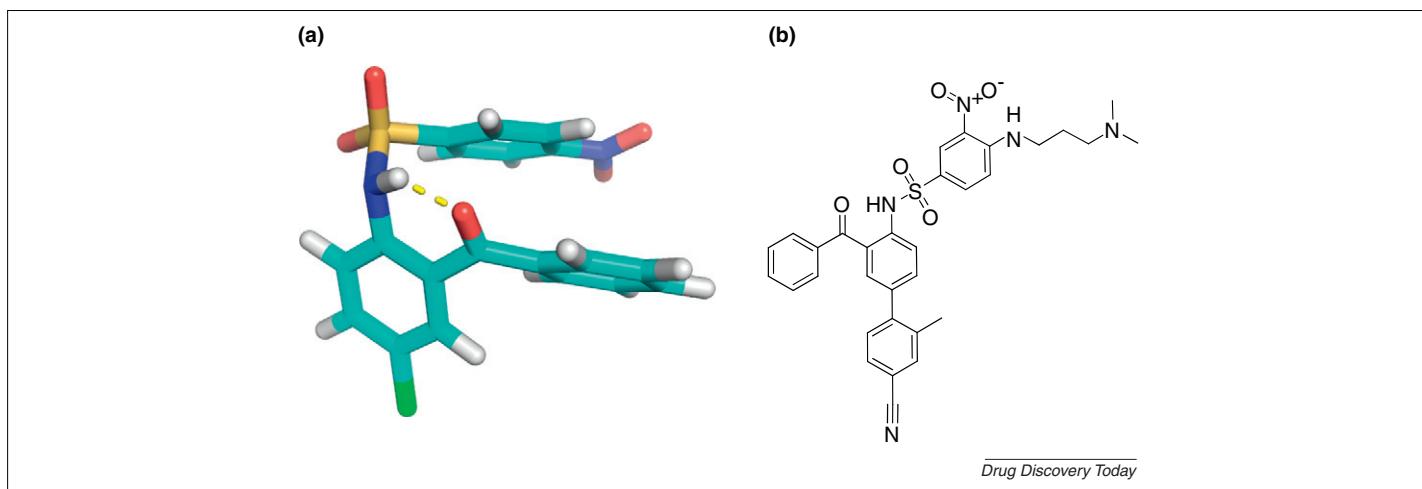
Software

The CSD is not just a crystal structure database of small molecules; it also has a powerful search engine and tools for query construction and structure mapping. The CSD package is a suitable topology hopping tool, by enabling users to define different topological requirements, including dihedral angle, point-to-plane distance and plane-to-plane angle [66]. ROCS is another powerful topology-based scaffold-hopping tool [108]. The grid-based method SHOP has also been used to identify novel scaffolds with significant different chemotypes from the queries they were derived from [111,112].

Concluding remarks

The pharmaceutical industry is facing dramatic challenges, primarily caused by reduced output of new medicines, drug price

pressures and global economic downturn [113]. Additionally, the unmet medical needs for rare and neglected diseases are not adequately addressed [114]. The situation strongly calls for more efficient ways to accelerate the pace of drug discovery. With the explosion of drug discovery related data in recent years [115–121], the mission becomes possible by making the best use of the available information. More recently, the National Institutes of Health Chemical Genomics Center (NCGC) prepared a complete collection of all approved drugs – the NCGC Pharmaceutical Collection (NPC) [122], and the NPC is being screened against multiple drug targets in a quantitative high-throughput screening (qHTS) format [123]. The high quality titration-based assay results will supply sufficient templates for scaffold-hopping approaches. Scaffold hopping, already widely accepted by medicinal chemists as a new design idea generator, is a proven tool

**FIGURE 15**

(a) The hit scaffold resulting from VS of crystal structures in CSD and (b) the structure of the new BCL-xL inhibitor. Abbreviations: BCL-xL: B-cell lymphoma-extra large; CSD: Cambridge Structural Database.

TABLE 1**The four types of scaffold-hopping methods, their pros and cons, and frequently used software for each method**

Category	Definition	Pros and cons	Software [Refs]
1°	 Heterocycle replacement	Pros: <ul style="list-style-type: none"> (1) High success rate (2) Immediate design Cons: <ul style="list-style-type: none"> (1) IP position (2) Limited changes in properties 	MORPH [45] and Recore [48]
2°	 Ring opening closure	Pros: <ul style="list-style-type: none"> (1) Improve binding (2) Improve stability Cons: <ul style="list-style-type: none"> (1) Reduce solubility (2) Flatten molecule (3) Synthetic feasibility 	CSD [67]
3°	 Pseudopeptide peptidomimetic	Pros: <ul style="list-style-type: none"> Ready templates from bioactive peptides or protein–protein interactions Cons: <ul style="list-style-type: none"> Metabolic stability is a concern, especially for pseudopeptides 	Recore [48], CAVEAT [87] and pharmacophore modeling tools from CCG [100], Accelrys [101] and Schrodinger [102]
4°	 Topology-based hopping	Pros: <ul style="list-style-type: none"> Significantly different scaffold, implying novel properties Cons: <ul style="list-style-type: none"> Lower success rate 	CSD [67], ROCS [108] and SHOP [112,124]

useful for overcoming undesirable properties, such as poor exposure, toxicity, or unfavorable IP position [16]. This article summarizes strategies commonly used in scaffolding hopping, and it classifies these methods into four basic categories (Table 1). Successful stories are highlighted for each category, while the limitations are also analyzed. At present, scaffold-hopping efforts are largely unsupervised and focus on novelty of derivative compounds. Future software developments in scaffold hopping also need to incorporate guiding elements for achieving acceptable pharmacological properties, such as improved absorption, better PK profiles and reduced toxicity. This would enable for a generalized scaffold-hopping approach that

maximizes the chemical diversity while maintaining or even improving the biological activity and pharmacological profiles of the original drug.

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References

- Russ, A.P. and Lampel, S. (2005) The druggable genome: an update. *Drug Discov. Today* 10, 1607–1610
- Hopkins, A.L. and Groom, C.R. (2002) The druggable genome. *Nat. Rev. Drug Discov.* 1, 727–730
- Boehm, H.J. et al. (2004) Scaffold hopping. *Drug Discov. Today: Technol.* 1, 217–224
- Sneader, W. (1996) *Drug Prototypes and Their Exploitation*. Wiley
- Martin, Y.C. and Muchmore, S. (2009) Beyond QSAR: lead hopping to different structures. *QSAR Comb. Sci.* 28, 797–801
- Cramer, R.D. et al. (2004) 'Lead hopping'. Validation of topomer similarity as a superior predictor of similar biological activities. *J. Med. Chem.* 47, 6777–6791
- Schneider, G. et al. (2006) Scaffold-hopping: how far can you jump? *QSAR Comb. Sci.* 25, 1162–1171
- Cramer, R.D. et al. (1999) Prospective identification of biologically active structures by topomer shape similarity searching. *J. Med. Chem.* 42, 3919–3933
- Schneider, G. et al. (1999) 'Scaffold-Hopping' by topological pharmacophore search: a contribution to virtual screening. *Angew. Chem. Int. Ed.* 38, 2894–2896
- Brown, N. and Jacoby, E. (2006) On scaffolds and hopping in medicinal chemistry. *Mini Rev. Med. Chem.* 6, 1217–1229
- Mauser, H. and Guba, W. (2008) Recent developments in de novo design and scaffold hopping. *Curr. Opin. Drug Discov. Dev.* 11, 365–374
- Jenkins, J.L. et al. (2004) A 3D similarity method for scaffold hopping from known drugs or natural ligands to new chemotypes. *J. Med. Chem.* 47, 6144–6159
- Stiefl, N. et al. (2006) Erg: 2D pharmacophore descriptions for scaffold hopping. *J. Chem. Inf. Model.* 46, 208–220
- Bohl, M. et al. (2006) Unsupervised 3D ring template searching as an ideas generator for scaffold hopping: use of the LAMDA, RigFit, and field-based similarity search (FBSS) methods. *J. Chem. Inf. Model.* 46, 1882–1890
- Gardiner, E.J. et al. (2011) Effectiveness of 2D fingerprints for scaffold hopping. *Future Med. Chem.* 3, 405–414
- Brown, N. et al. (2010) Bioisosteric replacement and scaffold hopping in lead generation and optimization. *Mol. Inform.* 29, 366–385
- Renner, S. and Schneider, G. (2006) Scaffold-hopping potential of ligand-based similarity concepts. *ChemMedChem* 1, 181–185
- Zhang, Q. and Muegge, I. (2006) Scaffold hopping through virtual screening using 2D and 3D similarity descriptors: ranking, voting, and consensus scoring. *J. Med. Chem.* 49, 1536–1548
- Martin, Y.C. et al. (2002) Do structurally similar molecules have similar biological activity? *J. Med. Chem.* 45, 4350–4358
- Wassermann, A.M. and Bajorath, J. (2010) Chemical substitutions that introduce activity cliffs across different compound classes and biological targets. *J. Chem. Inf. Model.* 50, 1248–1256
- Labute, P. et al. (2001) Flexible alignment of small molecules. *J. Med. Chem.* 44, 1483–1490
- White, J.M. and Rumbold, G.R. (1988) Behavioural effects of histamine and its antagonists: a review. *Psychopharmacology (Berl.)* 95, 1–14
- Soter, N.A. (1990) Urticaria: current therapy. *J. Allergy Clin. Immunol.* 86 (Pt 2), 1009–1014
- Mylecharane, E.J. (1991) 5-HT2 receptor antagonists and migraine therapy. *J. Neurol.* 238 (Suppl. 1), S45–S52
- Piwiński, J.J. et al. (1991) Dual antagonists of platelet activating factor and histamine. Identification of structural requirements for dual activity of N-acyl-4-(5,6-dihydro-11H-benzo[5,6]cyclohepta-[1,2-b]pyridin-11-ylidene)piperidines. *J. Med. Chem.* 34, 457–461
- Southall, N.T. and Ajay, (2006) Kinase patent space visualization using chemical replacements. *J. Med. Chem.* 49, 2103–2109
- FitzGerald, G.A. (2003) COX-2 and beyond: approaches to prostaglandin inhibition in human disease. *Nat. Rev. Drug Discov.* 2, 879–890
- Bostroem, J. et al. (2007) Scaffold hopping, synthesis and structure–activity relationships of 5,6-diaryl-pyrazine-2-amide derivatives: a novel series of CB1 receptor antagonists. *Bioorg. Med. Chem.* 15, 4077–4084
- Berggren, A.I.K. et al. (2004) Preparation of 4,5-diarylthiazole derivatives as CB1 ligands. Astrazeneca Ab/Astrazeneca UK Limited 34 pp.
- Berggren, A.I.K. et al. (2004) Astrazeneca Ab/Astrazeneca UK Limited 42 pp.
- Nicholls, A. and Grant, J.A. (2005) Molecular shape and electrostatics in the encoding of relevant chemical information. *J. Comput. Aided Mol. Des.* 19, 661–686
- Szivinska, I. et al. (2008) Cannabinoid receptors 1 and 2 (CB1 and CB2), their distribution, ligands and functional involvement in nervous system structures – a short review. *Pharmacol. Biochem. Behav.* 90, 501–511
- Lavey, B.J. et al. (2007) Optimization of triaryl bis-sulfones as cannabinoid-2 receptor ligands. *Bioorg. Med. Chem. Lett.* 17, 3760–3764
- Ritchie, T.J. and Macdonald, S.J. (2009) The impact of aromatic ring count on compound developability – are too many aromatic rings a liability in drug design? *Drug Discov. Today* 14, 1011–1020
- Ishikawa, M. and Hashimoto, Y. (2011) Improvement in aqueous solubility in small molecule drug discovery programs by disruption of molecular planarity and symmetry. *J. Med. Chem.* 54, 1539–1554
- Sun, H. (2004) A universal molecular descriptor system for prediction of log *P*, log *S*, logBB, and absorption. *J. Chem. Inf. Comput. Sci.* 44, 748–757
- Zarghi, A. et al. (2007) Design and synthesis of new rofecoxib analogs as selective cyclooxygenase-2 (COX-2) inhibitors: replacement of the methanesulfonyl pharmacophore by a *N*-acetylsulfonamido bioisostere. *J. Pharm. Pharm. Sci.* 10, 159–167
- Uddin, M.J. et al. (2003) Design and synthesis of novel celecoxib analogues as selective cyclooxygenase-2 (COX-2) inhibitors: replacement of the sulfonamide pharmacophore by a sulfonylazide bioisostere. *Bioorg. Med. Chem.* 11, 5273–5280
- DeWitt, D.L. (1999) Cox-2-selective inhibitors: the new super aspirins. *Mol. Pharmacol.* 55, 625–631
- Rimon, G. et al. (2010) Coxibs interfere with the action of aspirin by binding tightly to one monomer of cyclooxygenase-1. *Proc. Natl Acad. Sci. U. S. A.* 107, 28–33
- Wang, J.L. et al. (2010) The novel benzopyran class of selective cyclooxygenase-2 inhibitors. Part 2: The second clinical candidate having a shorter and favorable human half-life. *Bioorg. Med. Chem. Lett.* 20, 7159–7163
- Gierse, J.K. et al. (1996) A single amino acid difference between cyclooxygenase-1 (COX-1) and -2 (COX-2) reverses the selectivity of COX-2 specific inhibitors. *J. Biol. Chem.* 271, 15810–15814
- Gans, K.R. et al. (1990) Anti-inflammatory and safety profile of DuP 697, a novel orally effective prostaglandin synthesis inhibitor. *J. Pharmacol. Exp. Ther.* 254, 180–187
- Walker, M.C. et al. (2001) A three-step kinetic mechanism for selective inhibition of cyclo-oxygenase-2 by diarylheterocyclic inhibitors. *Biochem. J.* 357 (Pt 3), 709–718
- Beno, B.R. and Langley, D.R. (2010) MORPH: a new tool for ligand design. *J. Chem. Inf. Model.* 50, 1159–1164
- Pitt, W.R. et al. (2009) Heteroaromatic rings of the future. *J. Med. Chem.* 52, 2952–2963

REVIEWS

- 47 Lee, M.L. and Schneider, G. (2001) Scaffold architecture and pharmacophoric properties of natural products and trade drugs: application in the design of natural product-based combinatorial libraries. *J. Comb. Chem.* 3, 284–289
- 48 Maass, P. *et al.* (2007) Recore: a fast and versatile method for scaffold hopping based on small molecule crystal structure conformations. *J. Chem. Inf. Model.* 47, 390–399
- 49 Vieth, M. *et al.* (2004) Characteristic physical properties and structural fragments of marketed oral drugs. *J. Med. Chem.* 47, 224–232
- 50 Hall, A. *et al.* (2008) Discovery of a novel indole series of EP1 receptor antagonists by scaffold hopping. *Bioorg. Med. Chem. Lett.* 18, 2684–2690
- 51 Evelyn, C.R. *et al.* (2010) Design, synthesis and prostate cancer cell-based studies of analogs of the Rho/MKL1 transcriptional pathway inhibitor, CCG-1423. *Bioorg. Med. Chem. Lett.* 20, 665–672
- 52 Hodge, C.N. and Pierce, J. (1993) A diazine heterocyclic replaces a six-membered hydrogen-bonded array in the active site of scytalone dehydratase. *Bioorg. Med. Chem. Lett.* 3, 1605–1608
- 53 Eastwood, P. *et al.* (2010) Discovery of potent and selective bicyclic A(2B) adenosine receptor antagonists via bioisosteric amide replacement. *Bioorg. Med. Chem. Lett.* 20, 1634–1637
- 54 Panek, R.L. *et al.* (1997) *In vitro* pharmacological characterization of PD 166285, a new nanomolar potent and broadly active protein tyrosine kinase inhibitor. *J. Pharmacol. Exp. Ther.* 283, 1433–1444
- 55 Furet, P. *et al.* (2008) Entry into a new class of protein kinase inhibitors by pseudo ring design. *Bioorg. Med. Chem. Lett.* 18, 897–900
- 56 Furet, P. *et al.* (2003) Identification of a new chemical class of potent angiogenesis inhibitors based on conformational considerations and database searching. *Bioorg. Med. Chem. Lett.* 13, 2967–2971
- 57 Fujisaka, Y. *et al.* (2010) Phase 1 study of the investigational, oral angiogenesis inhibitor motesanib in Japanese patients with advanced solid tumors. *Cancer Chemother. Pharmacol.* 66, 935–943
- 58 Tasler, S. *et al.* (2009) N-substituted 2'-(*aminoaryl*)benzothiazoles as kinase inhibitors: hit identification and scaffold hopping. *Bioorg. Med. Chem. Lett.* 19, 1349–1356
- 59 Velicky, J. *et al.* (2010) Novel 3-aminopyrazole inhibitors of MK-2 discovered by scaffold hopping strategy. *Bioorg. Med. Chem. Lett.* 20, 1293–1297
- 60 Lovering, F. *et al.* (2009) Escape from flatland: increasing saturation as an approach to improving clinical success. *J. Med. Chem.* 52, 6752–6756
- 61 Reid, R.C. *et al.* (2004) Countering cooperative effects in protease inhibitors using constrained beta-strand-mimicking templates in focused combinatorial libraries. *J. Med. Chem.* 47, 1641–1651
- 62 Venkatraman, S. *et al.* (2005) Design and synthesis of depeptidized macrocyclic inhibitors of hepatitis C NS3-4A protease using structure-based drug design. *J. Med. Chem.* 48, 5088–5091
- 63 Pompei, M. *et al.* (2010) Novel P2–P4 macrocyclic inhibitors of HCV NS3/4A protease by P3 succinamide fragment depeptidization strategy. *Bioorg. Med. Chem. Lett.* 20, 168–174
- 64 Singh, E.K. *et al.* (2008) Conformationally based design of macrocycles as antitumor agents. *Curr. Opin. Drug Discov. Dev.* 11, 544–552
- 65 Uto, Y. *et al.* (2010) Novel spiroperidine-based stearoyl-CoA desaturase-1 inhibitors: Identification of 1'-[6-[5-(pyridin-3-ylmethyl)-1,3,4-oxadiazol-2-yl]pyridazin-3-yl]-5-(trifluoromethyl)-3,4-dihydrospiro[chromene-2,4'-piperidine]. *Bioorg. Med. Chem. Lett.* 20, 746–754
- 66 Allen, F.H. and Taylor, R. (2004) Research applications of the Cambridge Structural Database (CSD). *Chem. Soc. Rev.* 33, 463–475
- 67 Allen, F.H. (2002) The Cambridge Structural Database: a quarter of a million crystal structures and rising. *Acta Crystallogr. B* 58 (Pt 3/Pt 1), 380–388
- 68 Kuhn, B. *et al.* (2010) Intramolecular hydrogen bonding in medicinal chemistry. *J. Med. Chem.* 53, 2601–2611
- 69 Adessi, C. and Soto, C. (2002) Converting a peptide into a drug: strategies to improve stability and bioavailability. *Curr. Med. Chem.* 9, 963–978
- 70 Sun, H. *et al.* (2004) A predictive pharmacophore model of human melanocortin-4 receptor as derived from the solution structures of cyclic peptides. *Bioorg. Med. Chem. Lett.* 12, 2671–2677
- 71 Fry, D. and Sun, H. (2006) Utilizing peptide structures as keys for unlocking challenging targets. *Mini Rev. Med. Chem.* 6, 979–987
- 72 Vassilev, L.T. (2007) MDM2 inhibitors for cancer therapy. *Trends Mol. Med.* 13, 23–31
- 73 Vassilev, L.T. (2005) p53 Activation by small molecules: application in oncology. *J. Med. Chem.* 48, 4491–4499
- 74 Vassilev, L.T. *et al.* (2004) *In vivo* activation of the p53 pathway by small-molecule antagonists of MDM2. *Science* 303, 844–848
- 75 Chai, J. *et al.* (2000) Structural and biochemical basis of apoptotic activation by Smac/DIABLO. *Nature* 406, 855–862
- 76 Oosterom, J. *et al.* (1999) Conformation of the core sequence in melanocortin peptides directs selectivity for the melanocortin MC3 and MC4 receptors. *J. Biol. Chem.* 274, 16853–16860
- 77 Orner, B.P. *et al.* (2001) Toward proteomimetics: terphenyl derivatives as structural and functional mimics of extended regions of an alpha-helix. *J. Am. Chem. Soc.* 123, 5382–5383
- 78 Wist, A.D. *et al.* (2007) Structure–activity based study of the Smac-binding pocket within the BIR3 domain of XIAP. *Bioorg. Med. Chem.* 15, 2935–2943
- 79 Sebhate, I.K. *et al.* (2002) Design and pharmacology of N-[(3R)-1,2,3,4-tetrahydroisoquinolinium-3-ylcarbonyl]-*(1R*)-1-(4-chlorobenzyl)-2-[4-cyclohexyl-4-(1*H*-1,2,4-triazol-1-ylmethyl)piperidin-1-yl]-2-oxoethylamine (1), a potent, selective, melanocortin subtype-4 receptor agonist. *J. Med. Chem.* 45, 4589–4593
- 80 Vagner, J. *et al.* (2008) Peptidomimetics, a synthetic tool of drug discovery. *Curr. Opin. Chem. Biol.* 12, 292–296
- 81 Green, D.R. and Reed, J.C. (1998) Mitochondria and apoptosis. *Science* 281, 1309–1312
- 82 Charriaut-Marlangue, C. (2004) Apoptosis: a target for neuroprotection. *Therapie* 59, 185–190
- 83 Ferreira, C.G. *et al.* (2002) Apoptosis: target of cancer therapy. *Clin. Cancer Res.* 8, 2024–2034
- 84 Fleischer, A. *et al.* (2006) Modulating apoptosis as a target for effective therapy. *Mol. Immunol.* 43, 1065–1079
- 85 Yue, T.L. *et al.* (1999) Apoptosis: a potential target for discovering novel therapies for cardiovascular diseases. *Curr. Opin. Chem. Biol.* 3, 474–480
- 86 Cohen, F. *et al.* (2009) Orally bioavailable antagonists of inhibitor of apoptosis proteins based on an azabicyclooctane scaffold. *J. Med. Chem.* 52, 1723–1730
- 87 Lauri, G. and Bartlett, P.A. (1994) CAVEAT: a program to facilitate the design of organic molecules. *J. Comput. Aided Mol. Des.* 8, 51–66
- 88 Vučić, D. *et al.* (2005) Engineering ML-IAP to produce an extraordinarily potent caspase 9 inhibitor: implications for Smac-dependent anti-apoptotic activity of ML-IAP. *Biochem. J.* 385 (Pt 1), 11–20
- 89 Lawandi, J. *et al.* (2009) Constrained peptidomimetics reveal detailed geometric requirements of covalent prolyl oligopeptidase inhibitors. *J. Med. Chem.* 52, 6672–6684
- 90 Kaschina, E. and Unger, T. (2003) Angiotensin AT1/AT2 receptors: regulation, signalling and function. *Blood Press.* 12, 70–88
- 91 de Gasparo, M. *et al.* (2000) International union of pharmacology. XXIII. The angiotensin II receptors. *Pharmacol. Rev.* 52, 415–472
- 92 Garcia, K.C. *et al.* (1992) Three-dimensional structure of an angiotensin II-Fab complex at 3 Å: hormone recognition by an anti-idiotypic antibody. *Science* 257, 502–507
- 93 Nikiforovich, G.V. and Marshall, G.R. (1993) Three-dimensional recognition requirements for angiotensin agonists: a novel solution for an old problem. *Biochem. Biophys. Res. Commun.* 195, 222–228
- 94 Nikiforovich, G.V. *et al.* (1994) Conformational analysis of two cyclic analogs of angiotensin: implications for the biologically active conformation. *Biochemistry* 33, 3591–3598
- 95 Rosenstrom, U. *et al.* (2004) A selective AT2 receptor ligand with a gamma-turn-like mimetic replacing the amino acid residues 4–5 of angiotensin II. *J. Med. Chem.* 47, 859–870
- 96 Schmidt, B. *et al.* (1997) Design, synthesis, and biological activities of four angiotensin II receptor ligands with gamma-turn mimetics replacing amino acid residues 3–5. *J. Med. Chem.* 40, 903–919
- 97 Rosenstrom, U. *et al.* (2006) Design, synthesis, and incorporation of a beta-turn mimetic in angiotensin II forming novel pseudopeptides with affinity for AT1 and AT2 receptors. *J. Med. Chem.* 49, 6133–6137
- 98 Sanudo, M. *et al.* (2009) Synthesis of benzodiazepine beta-turn mimetics by an Ugi 4CC/Staudinger/aza-Wittig sequence. Solving the conformational behavior of the Ugi 4CC adducts. *J. Org. Chem.* 74, 2189–2192
- 99 Georgsson, J. *et al.* (2005) Angiotensin II pseudopeptides containing 1,3,5-trisubstituted benzene scaffolds with high AT2 receptor affinity. *J. Med. Chem.* 48, 6620–6631
- 100 <http://www.chemcomp.com/> (accessed October 2011)
- 101 <http://accelrys.com/> (accessed October 2011)
- 102 <http://www.schrodinger.com/> (accessed October 2011)
- 103 Sun, H. and Fry, D. (2007) Molecular modeling of melanocortin receptors. *Curr. Top. Med. Chem.* 7, 1042–1051
- 104 Hessler, G. and Baringhaus, K.H. (2010) The scaffold hopping potential of pharmacophores. *Drug Discov. Today: Technol.* 7, E263–E269
- 105 Franke, L. *et al.* (2007) Identification of natural-product-derived inhibitors of 5-lipoxygenase activity by ligand-based virtual screening. *J. Med. Chem.* 50, 2640–2646

- 106 Lutkenhaus, J. and Addinall, S.G. (1997) Bacterial cell division and the Z ring. *Annu. Rev. Biochem.* 66, 93–116
- 107 Hale, C.A. and de Boer, P.A. (1997) Direct binding of FtsZ to ZipA, an essential component of the septal ring structure that mediates cell division in *E. coli*. *Cell* 88, 175–185
- 108 Rush, T.S., 3rd *et al.* (2005) A shape-based 3-D scaffold hopping method and its application to a bacterial protein–protein interaction. *J. Med. Chem.* 48, 1489–1495
- 109 Sun, H. (2010) *De novo* design of potent BCL-xL inhibitors by scaffold hopping and data mining. *7th Annual Cancer Drugs Research and Development Conference*
- 110 Lee, E.F. *et al.* (2007) Crystal structure of ABT-737 complexed with Bcl-xL: implications for selectivity of antagonists of the Bcl-2 family. *Cell Death Differ.* 14, 1711–1713
- 111 Fontaine, F. *et al.* (2009) SHOP: a method for structure-based fragment and scaffold hopping. *ChemMedChem* 4, 427–439
- 112 Bergmann, R. *et al.* (2007) SHOP: scaffold HOPping by GRID-based similarity searches. *J. Med. Chem.* 50, 2708–2717
- 113 Bennani, Y.L. (2011) Drug discovery in the next decade: innovation needed ASAP. *Drug Discov. Today* 16, 779–792
- 114 Kuehn, B.M. (2009) NIH program to boost pipeline for drugs to treat rare and neglected diseases. *JAMA* 302, 133–134
- 115 Irwin, J.J. and Shoichet, B.K. (2005) ZINC – a free database of commercially available compounds for virtual screening. *J. Chem. Inf. Model.* 45, 177–182
- 116 Chen, B. *et al.* (2009) PubChem as a source of polypharmacology. *J. Chem. Inf. Model.* 49, 2044–2055
- 117 Chen, B. and Wild, D.J. (2010) PubChem BioAssays as a data source for predictive models. *J. Mol. Graph. Model.* 28, 420–426
- 118 Wang, Y. *et al.* (2010) An overview of the PubChem BioAssay resource. *Nucleic Acids Res.* 38 (Database issue), D255–D266
- 119 Wishart, D.S. *et al.* (2006) DrugBank: a comprehensive resource for in silico drug discovery and exploration. *Nucleic Acids Res.* 34 (Database issue), D668–D672
- 120 Liu, T. *et al.* (2007) BindingDB: a web-accessible database of experimentally determined protein-ligand binding affinities. *Nucleic Acids Res.* 35 (Database issue), D198–D201
- 121 Gaulton, A. *et al.* (2011) ChEMBL; European Bioinformatics Institute (EBI). Cambridge <http://www.ebi.ac.uk/chembl/> (accessed October, 2011)
- 122 Huang, R. *et al.* (2011) The NCGC pharmaceutical collection: a comprehensive resource of clinically approved drugs enabling repurposing and chemical genomics. *Sci. Transl. Med.* 3 80ps16
- 123 Inglese, J. *et al.* (2006) Quantitative high-throughput screening: a titration-based approach that efficiently identifies biological activities in large chemical libraries. *Proc. Natl Acad. Sci. U. S. A.* 103, 11473–11478
- 124 Bergmann, R. *et al.* (2009) SHOP: receptor-based scaffold HOPping by GRID-based similarity searches. *J. Chem. Inf. Model.* 49, 658–669